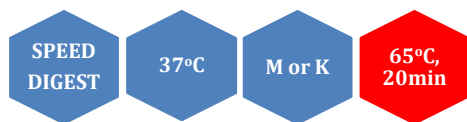


# Xba I



5' ...T▼CTAGA...3'  
3' ...AGATC▲T...5'

XbaI is a restriction enzyme purified from *Xanthomonas badrii*.

Catalogue No            143-1, 4000 U  
                                  143-2, 3x4000 U

Concentration            10-12u/μl and 40-60u/μl\*

\*Add an H to cat.# to order the high concentration

**Reagents supplied:** 10x M and 10x K buffer

**Unit substrate:** Lambda DNA (*dam*<sup>-</sup>/HindIII digest).

**Unit calculation assay conditions:** 50 mM NaCl, 10 mM Tris-HCl (pH 7.9 @ 25°C), 10 mM MgCl<sub>2</sub>, 1 mM dithiothreitol, 100 μg/ml BSA. Incubate at 37°C.

**Absence of contaminants:** 200 units of XbaI do not produce any unspecific cleavage products after 16 hrs incubation with 1 μg of λ DNAdam<sup>-</sup>/HindIII digest at 37°C. After 100-fold overdigestion with XbaI, greater than 98% of the DNA fragments can be ligated and recut with this enzyme.

**Storage buffer:** 100 mM KCl, 10 mM Tris-HCl (pH 7.4), 0.1 mM EDTA, 1 mM dithiothreitol, 500 μg/ml BSA and 50% glycerol. Store at -20°C.

**Heat inactivation:** 65°C for 20 minutes.

## Methylation Sensitivity:

dam methylation: Blocked by overlapping

dcm methylation: Not sensitive

CpG methylation: Not sensitive

## Percent Activity in MINOTECH Buffers

L	M	H	SH	A	K
50-75	100	75	75	75	100

## General reaction mixture:

10U XbaI	1μl
10x M or K buffer *	2μl
DNA substrate	<1μg
Sterile ultrapure water	Up to 20 μl

*Incubate for 15 min at 37°C*

\*In the case of M buffer we recommend the addition of BSA to a final concentration of 100 μg/ml.

## Frequency of Cutting

λ	Ad-2	Φx174	pUC18	M13mp18	pBR322
1	5	0	1	1	0



Lambda DNA (*dam*<sup>-</sup>) 0.7 % agarose