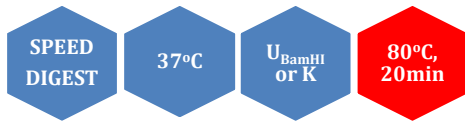


BamH I



5' ...G▼GATCC...3'
3' ...CCTAG▲G...5'

BamHI is a restriction enzyme purified from *Bacillus amyloliquefaciens* H.

Catalogue No 103-1, 3000 U
 103-2, 3x8000 U

Concentration 10-12u/μl and 40-60u/μl*

*Add an H to cat.# to order the high concentration

Reagents supplied: 10x U_{BamHI} and 10x K buffer.

Unit substrate: Lambda DNA.

Unit calculation assay conditions: 100 mM NaCl, 10 mM Tris-HCl (pH 7.9 @ 25°C), 5 mM MgCl₂, 1 mM dithiothreitol, 100 μg/ml BSA. Incubate at 37°C.

Absence of contaminants: 100 units of *BamHI* incubated for 16 hours at 37°C with 1 μg of λ DNA resulted in a DNA pattern free of detectable nuclease degradation as determined by agarose gel electrophoresis. After 50-fold overdigestion with *BamHI*, greater than 95% of the DNA fragments can be ligated and recut with this enzyme.

Storage buffer: 50 mM KCl, 10 mM Tris-HCl (pH 7.4), 0.1 mM EDTA, 1 mM dithiothreitol, 200 μg/ml BSA and 50% glycerol. Store at -20°C.

Heat inactivation: 80°C for 20 minutes.

Methylation Sensitivity:

dam methylation: Not sensitive
dcm methylation: Not sensitive
CpG methylation: Not sensitive

Star activity: Conditions of low ionic strength, high enzyme concentration, glycerol concentration >5% or pH >8.0 may result in star activity.

Percent Activity in MINOTECH Buffers

L	M	H	SH	A	K
75	75-100	100	50-75	75	100

General reaction mixture:

10U BamHI 1μl
10x U_{BamHI} or K buffer * 2μl
DNA substrate <1μg
Sterile ultrapure water Up to 20 μl
Incubate for 15 min at 37°C

*In the case of U_{BamHI} buffer we recommend the addition of BSA to a final concentration of 100 μg/ml.

Frequency of Cutting

λ	Ad-2	Φx174	pUC18	M13mp18	pBR322
5	3	0	1	1	1



Lambda DNA 0.7 % agarose